

# Nanotrap<sup>®</sup> Microbiome Combined; 10 mL Automated Protocol using NucleoMag<sup>®</sup> Kit and the KingFisher<sup>™</sup> Flex

**Objective:** This protocol uses Nanotrap Microbiome A Particles and Nanotrap Microbiome B Particles and Nanotrap Enhancement Reagent 3 to capture and concentrate microbes in environmental water samples. It is optimized for microbe capture from 10 mL samples and is compatible with MACHEREY-NAGEL NucleoMag DNA/RNA Water Kit. The automated script can process up to 24 samples at once and can be amended for the throughput in your lab.

## **Materials and equipment:**

Sample Type	
Environmental Water Samples	
Concentration Reagent	Vendor
Nanotrap Microbiome A Particles	Ceres Nanosciences; SKU# 44202
Nanotrap Microbiome B Particles	Ceres Nanosciences; SKU# 65202
Nanotrap Enhancement Reagent 3 (ER3) <sup>1</sup>	Ceres Nanosciences; SKU# 10113
Extraction Kit	Vendor
NucleoMag DNA/RNA Water Extraction Kit	MACHEREY-NAGEL; REF 744220.1
Materials/Equipment	Vendor
KingFisher™ Flex Purification System, KingFisher with 96 Deep-well Head	Thermo Fisher Scientific™; Cat# 5400630
KingFisher™ Flex 24 Deep Well head	Thermo Fisher Scientific™; Cat# 24074440
KingFisher™ Flex 24 Deep Well heating block	Thermo Fisher Scientific™; Cat# 24075440
KingFisher™ Flex 96 heating block	Thermo Fisher Scientific™; Cat# 24075420
KingFisher™ 24 deep-well plate (for Duo Prime, Flex and Presto)	Thermo Fisher Scientific™; Cat# 95040470
KingFisher™ 24 deep-well tip comb and plate (for Flex and Presto)	Thermo Fisher Scientific™; Cat# 97002610
KingFisher™ 96 deep-well plate, v-bottom, polypropylene (for Duo Prime, Flex and Presto)	Thermo Fisher Scientific™; Cat# 95040450
KingFisher™ 96 tip comb for deep-well magnets, 10 x 10 pcs/box (for Flex and Presto)	Thermo Fisher Scientific™; Cat# 97002534
General Reagents	Vendor
Molecular grade water	VWR; 45001-044

<sup>&</sup>lt;sup>1</sup> Precipitate can form in ER3 if stored below room temperature. Allow ER3 to return to room temperature to dissolve the precipitate (can invert 2-3 times to help resuspend, do not heat).

#### **Capture and Extract Microbes using Nanotrap Microbiome Particles**

#### Procedure:

### 1. Nanotrap Microbiome Combined NucleoMag KingFisher Flex Procedure-Part 1

- 1. Prepare "Sample Plate 1" and "Sample Plate 2"
  - 1. Invert environmental water sample 5 times to mix. After inverting, place on a flat surface for 45 seconds.
  - 2. Add 4,800 μL of environmental water sample to one well (one well per sample) of a new KingFisher 24 Well Deep Well Plate.
  - 3. Add another 4,800 µL of environmental water sample to the same well location on a second KingFisher™ 24 Well Deep Well Plate.
    - a) For example, if you loaded a sample into well A1 of the first plate, load the second volume of that sample into well A1 of the second plate.
  - 4. Add 50  $\mu$ L of Nanotrap Enhancement Reagent 3 (ER3) Solution to each sample on the two KingFisher 24 Well Deep Well sample plates (100  $\mu$ L total).
  - 5. Add 75 μL of Nanotrap<sup>®</sup> Microbiome A Particles to each sample on the two KingFisher 24 Well Deep Well sample plates (150 μL total).
  - 6. Add 75 μL of Nanotrap Microbiome B Particles to each sample on the two KingFisher 24 Well Deep Well sample plates (150 μL total).
- 2. Prepare "Lysis Plate"
  - 1. Add 500 μL of Lysis Buffer MWA1 to a new (the third) KingFisher 24 Well Deep Well Plate matching the number and location of the "Sample Plate" wells.
- 3. Prepare "Tip Plate"
  - 1. Insert a new tip comb into a new KingFisher 24 Well Deep Well Plate.
- 4. Run NT Script (Request file at sales @ceresnano.com)
  - 1. Run

#### NT Microbiome A B NucleoMag® 24 w heat Flex 10mL.bdz

- 2. Follow the on-screen instructions loading the previously prepared plates at the appropriate time.
- 5. Once the protocol is completed, the "Lysis Plate" will contain lysate that is ready to proceed to Part 2 (\*caution\* sample may be hot).

### 2. Nanotrap Microbiome Combined NucleoMag KingFisher Flex Procedure-Part 2

- 1. Prepare "NM Binding" Plate
  - 1. To a new KingFisher 96 Deep Well Plate, add 450  $\mu$ L of the cleared lysate (NT lysate) from each well of the lysis plate used in "Part 1 step 5" of the

- protocol. Keep track of which well contains which sample in this new bead binding plate.
- 2. Add 475 μL of Binding Buffer MWA2 to each well in which lysate was added.
- 3. Vortex the NucleoMag B-beads thoroughly and add 25 µL to each well.
  - a) Note: Binding mix (MWA2 + B-beads) can be pre-mixed before their addition to the plate.
- 2. Prepare "1st Wash MWA3" Plate
  - 1. Add 850 µL of Wash Buffer MWA3 to a new KingFisher 96 Deep Well Plate matching the number and location of the KingFisher 96 Deep Well Plate- "NM Binding" Plate wells.
- 3. Prepare "2<sup>nd</sup> Wash MWA3" Plate
  - 1. Add 850 µL of Wash Buffer MWA3 to a new KingFishe 96 Deep Well Plate matching the number and location of the KingFisher 96 Deep Well Plate- "NM Binding" Plate wells.
- 4. Prepare "3rd Wash MWA4" Plate
  - 1. Add 850 µL of Wash Buffer MWA4 to a new KingFisher 96 Deep Well Plate matching the number and location of the KingFisher 96 Deep Well Plate- "NM Binding" Plate wells.
- 5. Prepare "Elution" Plate
  - 1. Add 100 μL of Rnase-free water to a new KingFisher 96 Deep Well Plate matching the number and location of the KingFisher 96 Deep Well Plate-"NM Binding" Plate wells.
- 6. Prepare "Tip Plate"
  - Insert the KingFisher 96 Deep Well Comb into a new KingFisher 96 Deep Well Plate
- 7. Run Extraction Kit Script (Request file at sales @ceresnano.com)
  - 1. Run

#### NucleoMag\_DNA\_RNA\_Water\_CeresNanoTrap\_Flex\_Rev02.bdz

- 2. Follow the on-screen instructions loading the previously prepared plates at the appropriate time.
- 3. Once the protocol is completed, the KingFisher 96-Elution Plate contains eluates that are ready for downstream analysis or can be stored at -80°C.

Note: Multiple freeze-thaw cycles may cause degradation.

### Attachments: 2

KingFisher™ Flex

- 1. NT\_Microbiome\_A\_and\_B\_NucleoMag®\_24\_w\_heat\_Flex\_10mL.bdz
- 2. NucleoMag\_DNA\_RNA\_Water\_CeresNanoTrap\_Flex\_Rev02.bdz